Acarus Sample Types and Turnaround Times

Infectious Disease	Sample	Turnaround Time
Anaplasma phagocytophilum (qPCR)	0.5ml EDTA blood and/or tissue aspirates in EDTA tube and/or fresh tissue biopsy material*	3-5 working days
Babesia spp (qPCR)	0.5ml EDTA blood (or fresh tissue biopsy material* can be tested if available instead of blood e.g. splenic tissue)	3-5 working days
Bartonella henselae (qPCR)	0.5 ml EDTA blood and/or tissue aspirates in EDTA tube and/or fresh tissue biopsy material*	3-5 working days
Bartonella spp (qPCR)	0.5 ml EDTA blood and/or tissue aspirates in EDTA tube and/or fresh tissue biopsy material*	3-5 working days
Borrelia spp (Lyme disease) (PCR)	0.5ml EDTA blood (but this is not an ideal sample due to low sensitivity); 0.5ml synovial fluid in EDTA tube or skin biopsy from the tick bite site	3-5 working days
Ehrlichia/Anaplasma spp (PCR)	0.5ml EDTA blood and/or tissue aspirates in EDTA tube and/or fresh tissue biopsy material	3-5 working days
Heartworm screen (<i>Dirofilaria immitis</i> antigen and modified Knott's microfilariae test)	1ml EDTA blood AND 0.5ml serum	1-2 working days
Hepatozoon spp (PCR)	0.5ml EDTA blood (or fresh tissue biopsy material* can be tested if available instead of blood e.g. splenic tissue)	3-5 working days
Leishmania (qPCR)	0.5 ml EDTA blood and/or lymph node aspirates in EDTA tube and/or splenic aspirates in EDTA tube and/or bone marrow aspirates/biopsy in EDTA tube and/or fresh tissue biopsy material* and/or conjunctival swabs **	3-5 working days
Leishmania Antibody	1 ml EDTA blood or 0.5ml serum	5-7 working days

Reception Hours

Mon-Fri 9am - 5pm

Contact Us

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E: labs@langfordvets.co.uk acarus-lab@bristol.ac.uk





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SNAP®4Dx® serology screen:		
D. immitis Ag		
E. canis Ab	0.5ml EDTA or serum	1-2 working
A. phagocytophilum /		days
A. platys Ab		
B. burgdoferi Ab		

^{*} Fresh biopsy material should be placed on saline soaked gauze material in a sterile container to keep moist and container should then be sealed. Formalin fixed specimens are not recommended in the first instance.

^{**} Conjunctival swabs should be moistened with sterile saline before use. After collection, the end should be cut off and placed in a sterile container to keep moist and container should then be sealed.



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